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Data Sheet

PARP7 Chemiluminescent Assay Kit

Catalog #79729-2

DESCRIPTION: The *PARP7 Chemiluminescent Activity Assay Kit* is designed to measure PARP7 activity for screening and profiling applications. PARP7 is known to catalyze the NAD-dependent addition of poly (ADP-ribose) to histones. The PARP7 assay kit comes in a convenient 384-well format, with purified PARP7 enzyme, histone mixture and PARP7 assay buffer for 400 enzyme reactions. The key to the *PARP7 Chemiluminescent Activity Assay* is the biotinylated substrate. With this kit, only three simple steps are required for PARP7 reactions. First, histone proteins are coated on a 384-well plate. Next, the PARP7 biotinylated substrate is incubated with an assay buffer that contains the PARP7 enzyme. Finally, the plate is treated with streptavidin-HRP followed by addition of the ELISA ECL substrate to produce chemiluminescence that can then be measured using a chemiluminescence reader.

COMPONENTS:

Catalog #	Reagent	Amount	Storage	
80527	PARP7	2 x 20 µg	-80°C	***Avoid multiple freeze/ thaw cycles!***
52029	5x histone mixture	2 x 1 ml	-80°C	
79814	Opti-PARP 10x Assay Mixture Containing Biotinylated Substrate	2 x 300 µl	-80°C	
80602	10x PARP assay buffer	2 x 1 ml	-20°C	
79743	Blocking buffer 3	2 x 25 ml	+4°C	
80611	Streptavidin-HRP	2 x 100 µl	+4°C	
79670	ELISA ECL substrate A (translucent bottle)	2 x 6 ml	Room Temp.	
	ELISA ECL substrate B (brown bottle)	2 x 6 ml	Room Temp.	
	384-well plate	1	Room Temp.	

MATERIALS OR INSTRUMENTS REQUIRED BUT NOT SUPPLIED:
1x PBS buffer
PBST buffer (1x PBS, containing 0.05% Tween-20)
Luminometer or fluorescent microplate reader capable of reading chemiluminescence
Adjustable micropipettor and sterile tips
Rotating or rocker platform
DTT

APPLICATIONS: Great for studying enzyme kinetics and screening small molecular inhibitors for drug discovery and HTS applications.

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STABILITY: Up to 1 year when stored as recommended.

REFERENCE: Brown JA, Marala RB. *J. Pharmacol. Toxicol. Methods* 2002 **47:**137-41.

ASSAY PROTOCOL:
All samples and controls should be tested in duplicate.

Step 1: Coat histone solution onto a 384-well plate

1) Dilute **5x histone mixture** 1:5 with PBS.

2) Add 25 µl of diluted histone solution to each well and incubate at 4ºC overnight.

3) Discard histone solution and tap the plate onto clean paper towel to remove remaining solution.

4) Block the wells by adding 100 µl of **Blocking buffer 3** to every well. Incubate at room temperature for 60-90 minutes.

5) Tap plate onto clean paper towel to remove liquid.

6) Wash plate two times with 100 µl PBST buffer as described above.

**Note: Coated plates can be stored at 4ºC for 2-3 days before using. However, it is strongly recommended that a freshly coated plate be used whenever possible.*

Step 2: Ribosylation reaction

1) Prepare the master mixture: N wells x (1.25 µl **10x PARP buffer** + 1.25 µl **Opti-PARP 10X Assay mixture** + 7.5 µl distilled water + 2.5 µl 5 mM DTT). Add 12.5 µl to every well.

2) Prepare 10X concentrated inhibitor in an aqueous-based solution. *Note: Final DMSO concentration must be ≤1%. Higher DMSO levels can significantly decrease the enzyme activity. For example, to test an inhibitor dissolved in 100% DMSO at 10 µM, dilute 1 mM inhibitor with water to make a 100 µM inhibitor in 10% DMSO(aq). Then, add 2.5 µl of the 100 µM solution to the assay to make a 1% DMSO concentration in the final reaction mixture*

3) Add 2.5 µl of Inhibitor solution of each well labeled as “Test Inhibitor.” For the “Positive Control" and “Blank,” add 2.5 µl of the same solution without inhibitor (e.g. 10% DMSO (aq))

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	Positive Control	Test Inhibitor	Blank
10x PARP buffer	1.25 µl	1.25 µl	1.25 µl
10x Assay mixture	1.25 µl	1.25 µl	1.25 µl
Water	7.5 µl	7.5 µl	7.5 µl
5 mM DTT	2.5 µl	2.5 µl	2.5 µl
Test Inhibitor	–	2.5 µl	–
10% DMSO aq (Inhibitor buffer)	2.5 µl	–	2.5 µl
1x PARP buffer	–	–	10 µl
PARP7 (10 ng/µl)*	10 µl	10 µl	–
Total	25 µl	25 µl	25 µl

* If using a freshly coated plate, 100 ng/rxn of PARP7 gives good signal (S/B > 20). If using a plate that was coated and stored at 4ºC for one or more days, signal could become lower.

4) Prepare 1x PARP buffer by adding 1 part of **10x PARP buffer** to 9 parts distilled water (v/v) and add DTT to a final concentration of 1.25 mM.

5) To the wells designated as "Blank," add 10 µl of 1X PARP buffer.

6) Thaw **PARP7 enzyme** on ice. Upon first thaw, briefly spin tube containing enzyme to recover full content of the tube. Calculate the amount of **PARP7** required for the assay and dilute enzyme to 10 ng/µl with 1x PARP buffer*. Aliquot remaining **PARP7 enzyme** into single use aliquots. Store remaining undiluted enzyme in aliquots at -80°C. *Note: PARP7 enzyme is very sensitive to freeze/thaw cycles. Avoid multiple freeze/thaw cycles. Do not re-use thawed aliquots or diluted enzyme.*

7) Initiate reaction by adding 10 µl of diluted **PARP7 enzyme** to the wells designated “Positive Control” and "Test Inhibitor." Centrifuge the plate at 1000 rpm for 30 seconds and incubate at room temperature for 1 hour.

8) Discard the reaction mixture after 1 hour, and wash plate two times with 100 µl PBST buffer and tap plate onto clean paper towel as described above.

Step 3: Detection

1) Dilute **Streptavidin-HRP** 1:50 in **Blocking buffer 3.**

2) Add 25 µl of diluted Streptavidin-HRP to each well. Incubate for 30 min. at room temperature.

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3) Wash three times with 100 µl PBST buffer and tap plate onto clean paper towel as above as described above.

4) Just before use, mix on ice 25 µl **ELISA ECL substrate A** and 25 µl **ELISA ECL substrate B** and add 50 µl per well.

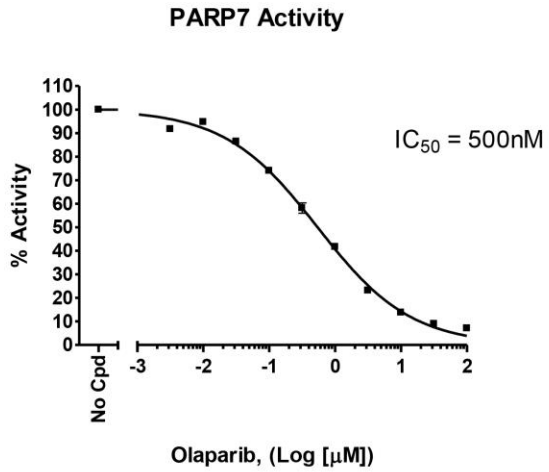
5) Immediately read the plate in a luminometer or microtiter-plate reader capable of reading chemiluminescence. The "Blank" value is subtracted from all other values.

Reading Chemiluminescence:

Chemiluminescence is the emission of light (luminescence) which results from a chemical reaction. The detection of chemiluminescence requires no wavelength selection because the method used is emission photometry and is not emission spectrophotometry.

To properly read chemiluminescence, make sure the plate reader is set for LUMINESCENCE mode. Typical integration time is 1 second, delay after plate movement is 100 msec. Do not use a filter when measuring light emission. Typical settings for the Synergy 2 BioTek plate reader are: use the "hole" position on the filter wheel; Optics position: Top; Read type: endpoint. Sensitivity may be adjusted based on the luminescence of a control assay without enzyme (typically we set this value as 100).

Example of Assay Results:



Inhibition of PARP7 activity by Olaparib, measured using the *PARP7 Chemiluminescent Activity Assay Kit,* (BPS Bioscience #79729). Luminescence was measured using a Bio-Tek microplate reader. *Data shown is lot-specific. For lot-specific information, please contact BPS Bioscience, Inc. at info@bpsbioscience.com*

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RELATED PRODUCTS:

Product Name	Catalog #	Size
PARP7 Enzyme	80507	10 µg
PARP7Chemiluminescent Assay Kit	80557	96 rxns.
PARP1 Assay Kit	80551	96 rxns.
PARP2 Assay Kit	80552	96 rxns.
PARP3 Assay Kit	80553	96 rxns.
PARP5A (TNKS1) Assay Kit	80573	96 rxns.
PARP5B (TNKS2) Assay Kit	80579	96 rxns.
PARP6 Assay Kit	80556	32 rxns.
PARP1 Enzyme	80501	10 µg
PARP2 Enzyme	80502	10 µg
PARP3 Enzyme	80503	10 µg
PARP6 Enzyme	80506	10 µg
TNKS2 (PARP5A) Enzyme	80504	10 µg
TNKS2 (PARP5B/C) Enzyme	80505	10 µg
PARP9 Enzyme	80509	10 µg
PARP11 Enzyme	80511	10 µg
PARP12 Enzyme	80512	10 µg

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TROUBLESHOOTING GUIDE

Problem	Possible Cause	Solution
Luminescence signal of positive control reaction is weak	PARP7 enzyme has lost activity	Enzyme loses activity upon repeated freeze/thaw cycles. Use fresh enzyme (PARP7, BPS Bioscience #80527). Store enzyme in single-use aliquots. Increase time of enzyme incubation. Increase enzyme concentration.
	Incorrect settings on instruments	Refer to instrument instructions for settings to increase sensitivity of light detection. See section on "Reading Chemiluminescence" above.
	Chemiluminescent reagents mixed too soon	Chemiluminescent solution should be used within 15 minutes of mixing. Ensure both reagents are properly mixed.
Luminescent signal is erratic or varies widely among wells	Inaccurate pipetting/technique	Run duplicates of all reactions. Use a multichannel pipettor. Use master mixes to minimize errors.
	Bubbles in wells	Pipette slowly to avoid bubble formation. Tap plate lightly to disperse bubbles; be careful not to splash between wells.
Background (signal to noise ratio) is high	Insufficient washes	Be sure to include blocking steps after wash steps. Increase number of washes. Increase wash volume. Increase Tween-20 concentration to 0.1% in TBST.
	Sample solvent is inhibiting the enzyme	Run negative control assay including solvent. Maintain DMSO level at <1% Increase time of enzyme incubation.
	Results are outside the linear range of the assay	Use different concentrations of enzyme (PARP7, BPS Bioscience #80527) to create a standard curve.

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