

MURF3 Intrachain TR-FRET Assay Kit

Description

The MURF3 intrachain TR-FRET Assay Kit is a sensitive high-throughput screening (HTS) TR-FRET Assay Kit, designed to measure MURF3 auto-ubiquitination activity in a homogeneous 384 reaction format. It utilizes a Europium cryptate-labeled Ub (donor) as well as Cy5-labeled Ub (acceptor) to complete the TR-FRET pairing. Since both the TR-FRET donor and acceptor are incorporated into poly-ubiquitin chains formed on MURF3, this FRET-based assay requires no time-consuming washing steps, making it especially suitable for HTS applications as well as real-time kinetic analyses of polyubiquitination.

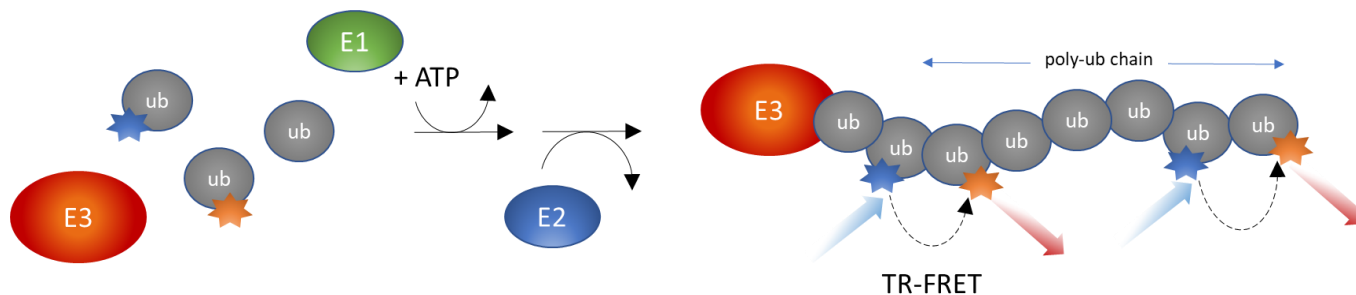


Figure 1: MURF3 intrachain TR-FRET Assay Kit schematic

Background

Covalent conjugation to ubiquitin (Ub) is one of the major post-translational modifications that regulates protein stability, function, and localization. Ubiquitination is the concerted action of three enzymes: a Ub-activating enzyme (E1), a Ub-conjugating enzyme (E2), and a Ub ligase (E3). The specificity and efficiency of ubiquitination are largely determined by the E3 enzyme, which directs the last step of the Ub-conjugating cascade by binding to both an E2~Ub conjugate and a substrate protein. This step ensures the transfer of Ub from E2~Ub to the substrate, leading to its mono- or poly-ubiquitination.

Muscle-Specific RING Finger Protein 3 (MURF3, also known as TRIM54) is a protein that associates with microtubules in cardiac and skeletal muscle cells, where it is involved in myoblast differentiation and microtubule network development. MURF3 is known to interact with and ubiquitinate filamin, which plays critical roles in actin filament formation and cytoskeletal structure. MURF3 mutations are associated with myopathies and cardiomyopathies, and therefore MURF3 is a potential target for the treatment of these diseases.

Applications

- Screen molecules that inhibit MURF3 Ub ligase activity in HTS applications
- Determine compound IC₅₀
- Perform MURF3 real-time kinetic analyses.

Supplied Materials

Catalog #	Name	Amount	Storage	
80301	UBE1 (UBA1), FLAG-tag (E1)*	50 µg	-80°C	Avoid multiple freeze/thaw cycles
80314	UbcH5b, His-Tag (E2)*	60 µg	-80°C	
81054	MURF3, SUMO-His-tags (E3)*	10 µg	-80°C	
78307	TRF Ubiquitin Mix (200x)	50 µl	-80°C	
	ATP (4 mM)	2 x 1 ml	-80°C	
	U2 Assay Buffer	2 x 10 ml	-80°C	
	White, nonbinding Corning, low volume microtiter plate		Room Temp	

* The initial concentration of enzyme is lot-specific and will be indicated on the tube containing the protein.

Materials Required but Not Supplied

- Fluorescent microplate reader capable of measuring Time Resolved Fluorescence Resonance Energy Transfer (TR-FRET)
- Adjustable micropipettor and sterile tips
- Rotating or rocker platform

Storage Conditions

This assay kit will perform optimally for up to 6 months from date of receipt when the materials are stored as directed. **Avoid multiple freeze/thaw cycles!**

Safety

This product is for research purposes only and not for human or therapeutic use. This product should be considered hazardous and is harmful by inhalation, in contact with skin, eyes, clothing, and if swallowed. If contact occurs, wash thoroughly.

Contraindications

The MURF3 intrachain TR-FRET Assay Kit is compatible with up to 1% final DMSO concentration. We recommend preparing the inhibitor in no higher than 5% DMSO solution in buffer and using 4 µl per well.

Assay Protocol

- All samples and controls should be performed in triplicate
 - The assay should include a “Blank”, a “Positive control”, and a “Negative control”
 - Calculate each protein, assay buffer, and ATP into single-use aliquots for the desired number of reactions per assay test.
1. Thaw **UBE1, UBCH5b, MURF3, TRF Ubiquitin Mix, U2 Assay Buffer, and ATP** on ice. Briefly spin the tubes to recover their full contents.
 2. Prepare 5-fold concentrated TRF Ubiquitin Mix in U2 Assay Buffer (prepare a 40-fold dilution of the provided 200x TRF Ubiquitin Mix)

3. Calculate the amount of protein required for the assay and prepare the appropriate amounts of diluted proteins. The concentration of each protein is lot-specific and is indicated on the tube. Verify the initial concentration and dilute accordingly.
 - a. Dilute UBE1 in U2 Assay Buffer at 96 ng/μl (800 nM - the final concentration in the reaction is 40 nM);
 - b. Dilute UBCH5b in U2 Assay Buffer at 144 ng/μl (2 μM - the final concentration in the reaction is 100 nM);
 - c. Dilute MURF3 in U2 Assay Buffer at 4.03 ng/μl (100 nM - the final concentration in the reaction is 25 nM);

Note: Keep all diluted proteins on ice until use. Do not freeze and re-use the diluted proteins.

Note: Aliquot the remaining unused, undiluted protein into 2-4 aliquots as may be necessary (single use aliquots) and store them at -80°C. Aliquot assay buffer and ATP and store at -80°C.

4. Prepare the Test Inhibitor (4 μl/well): for a titration, prepare serial dilutions at concentrations 5-fold higher than the desired final concentrations. The final volume of the reaction is 20 μl.

Without DMSO

- a. If the Test Inhibitor is soluble in water, prepare a solution of the compound in U2 Assay Buffer that is 5-fold higher than the final desired concentration.

Or

With DMSO

- a. If the Test Inhibitor is dissolved in DMSO, prepare a solution of the compound in DMSO that is 100-fold higher than the highest concentration of the serial dilution. Then dilute 20-fold in Assay Buffer 2 (at this step the compound concentration is 5-fold higher than the desired final concentration). The concentration of DMSO in the dilution is now 5%.
 - b. Prepare serial dilutions of the Test Inhibitor at concentrations 5-fold higher than the desired final concentrations using 5% DMSO in U2 Assay Buffer to keep the concentration of DMSO constant.
 - c. For positive and negative controls, prepare 5% DMSO in U2 Assay Buffer (vol/vol) so that all wells contain the same amount of DMSO (Diluent Solution).
5. To the wells designated as "Blank", add 4 μl of **5x TRF Ubiquitin Mix** + 1 μl of **UBE1** + 1 μl of **UBCH5b** + 4 μl of **diluent solution** (for example DMSO 5%) + 5 μl of **U2 Assay Buffer**.

Component	Blank
TRF Ubiquitin Mix (5x)	4 μ l
UBE1	1 μ l
UBCH5b	1 μ l
MURF3	-
Test Compound	-
Diluent solution* (no inhibitor)	4 μ l
U2 Assay Buffer	5 μ l
ATP (4 mM)	5 μ l
Total	20 μl

*The diluent solution contains the assay buffer with the same concentration of solvent (e.g. DMSO) as the test compound solution.

- Prepare a Master Mix using diluted reagents: N wells \times (4 μ l of **5x TRF Ubiquitin Mix** + 1 μ l of **UBE1** + 1 μ l of **UBCH5b** + 5 μ l of **MURF3**).
- Add 11 μ l of Master Mix to each well designated for the “Negative Control”, “Positive Control”, “Test Inhibitor”.
- Add 4 μ l of Test Inhibitor to each well designated “Test Inhibitor”. For “Positive Control” and “Negative Control”, add 4 μ l of the diluent solution without inhibitor.
- Initiate the reaction by adding 5 μ l of **ATP** to the wells labeled “Positive Control,” “Test Inhibitor,” and “Blank.” Add 5 μ l of **U2 Assay Buffer** to the well designated “Negative Control.” Cover the plate with a plate sealer. Incubate the reaction at room temperature for two hours or at 30°C for one hour.

Component	Test Inhibitor	Negative Control	Positive Control
Master Mix	11 μ l	11 μ l	11 μ l
Test compound	4 μ l	-	-
Diluent solution* (no inhibitor)	-	4 μ l	4 μ l
U2 Assay Buffer	-	5 μ l	-
ATP (4 mM)	5 μ l	-	5 μ l
Total	20 μl	20 μl	20 μl

*The diluent solution contains the assay buffer with the same concentration of solvent (e.g. DMSO) as the test compound solution.

- Read the fluorescent intensity in a microtiter-plate reader capable of measuring TR-FRET.

Note: Two sequential measurements should be conducted. Eu-donor emission should be measured at 620 nm followed by dye-acceptor emission at 665 nm. Data analysis is performed using the TR-FRET ratio (665 nm emission/620 nm emission). “Blank” value is subtracted from all other values.

Instrument Settings

Eu-donor emission		Dye-acceptor emission	
Reading Mode	Time Resolved	Reading Mode	Time Resolved
Excitation Wavelength	317±20 nm	Excitation Wavelength	317±20 nm
Emission Wavelength	620±10 nm	Emission Wavelength	665±10 nm
Lag Time	60 µs	Lag Time	60 µs
Integration Time	500 µs	Integration Time	500 µs

CALCULATING RESULTS:

Data analysis is performed using the TR-FRET ratio (665 nm emission/620 nm emission). “Blank” value is subtracted from all other values.

When percentage activity is calculated, the FRET value from the Blank (it is expected that Blank and Negative Control represent similar value) can be set as zero percent activity and the FRET value from the positive control can be set as one hundred percent activity.

$$\% \text{ Activity} = \frac{\text{FRET}_s - \text{FRET}_{\text{neg}}}{\text{FRET}_p - \text{FRET}_{\text{neg}}} \times 100\%$$

Where FRET_s = Sample FRET, FRET_{blank} = Blank FRET, and FRET_p = Positive control FRET.

Example Results

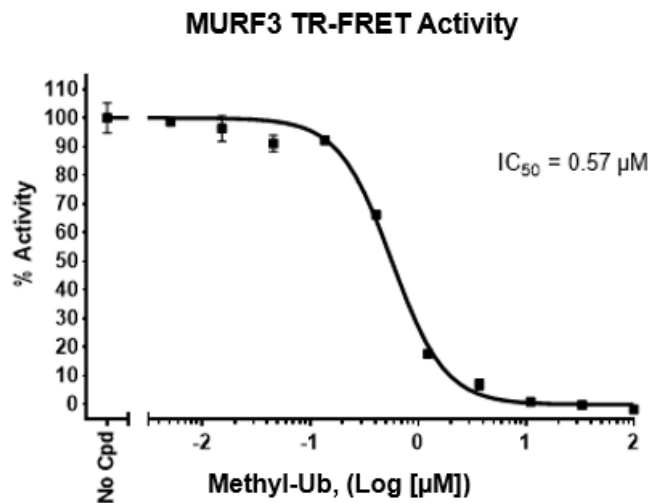


Figure 2: MURF3 TR-FRET Activity.

Inhibition of MURF3 auto-ubiquitination by Methylated Ubiquitin, measured using the MURF3 intrachain TR-FRET Assay Kit (BPS Bioscience #78555). Data shown is representative. For lot-specific information, please contact BPS Bioscience, Inc. at support@bpsbioscience.com.

Troubleshooting Guide

Visit bpsbioscience.com/assay-kits-faq for detailed troubleshooting instructions. For all further questions, please email support@bpsbioscience.com

Related Products

Products	Catalog #	Size
Cereblon intrachain TR-FRET Assay Kit	78301	384 reactions
Cereblon Ubiquitination Homogenous Assay Kit	79881	384 reactions
MDM2 intrachain TR-FRET Assay Kit	78302	384 reactions
SMURF1 intrachain TR-FRET Assay Kit	78303	384 reactions
VHL intrachain TR-FRET Assay Kit	78305	384 reactions
XIAP intrachain TR-FRET Assay Kit	78306	384 reactions
MDM2 TR-FRET Assay Kit	79773	384 reactions
CBL-B TR-FRET Assay Kit	79575	384 reactions
c-CBL TR-FRET Assay Kit	79786	384 reactions
UBCH13 TR-FRET Assay Kit	79741	384 reactions
UBCH5a TR-FRET Assay Kit	79900	384 reactions
UBCH5c TR-FRET Assay Kit	79901	384 reactions
UBCH5b TR-FRET Assay Kit	79896	384 reactions