Description

The PCSK9(His)-LDLR Binding Assay Kit is an ELISA-type 96-well format assay designed for screening and profiling purposes. The Assay Kit contains His-tag PCSK9, purified LDLR ectodomain and Anti-His-tag Horseradish Peroxidase (HRP)-conjugated antibody in sufficient amount for 100 binding reactions. Moreover, two pre-formulated assay buffers are supplied to validate PCSK9-LDLR binding affinity in either neutral or acidic binding conditions. The assay takes advantage of the high sensitivity of detection of His-tag PCSK9 by Anti-His HRP-antibody. First, the LDLR ectodomain is coated on a 96-well plate. Next, PCSK9 is incubated with LDLR on the plate. Finally, the plate is treated with Anti-His HRP-antibody followed by addition of an HRP substrate to produce chemiluminescence, measured using a chemiluminescence reader.

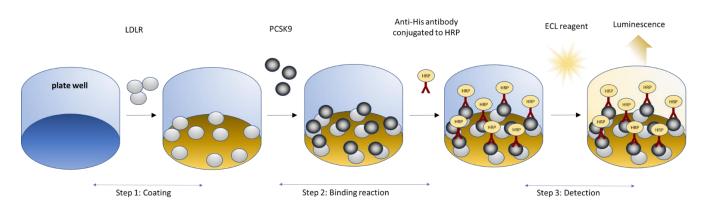


Figure 1: Illustration of the ELISA-based PCSK9(His)-LDLR Binding Assay Kit.

Background

PCSK9 (Proprotein convertase subtilisin/kexin type 9) functions as a negative regulator of hepatic low-density lipoprotein receptors (LDLRs) and therefore is a critical regulator of cholesterol metabolism. It is an endopeptidase that binds to the EGFR-like ectodomain of LDLR, leading to LDLR degradation, which in turn, results in increased circulating LDL. Inhibiting the PCSK9-LDLR interaction is an increasingly desirable therapeutic approach for lowering LDL-cholesterol levels to replace or supplement statins. New therapies are critical for addressing atherosclerosis, stroke, heart disease, and other cardiovascular disorders.

Applications

Screen small molecules and antibodies that inhibit the binding of PCSK9 to LDLR ectodomain.



Supplied Materials

Catalog #	Name	Amount	Storage		
71204	PCSK9(His)*	10 µg	-80°C	Avoid multiple	
71205	LDLR*	10 µg	-80°C		
	Anti-His HRP-D Antibody-D	10 µl	-80°C		
33298	3x PL-01 Assay Buffer	50 ml	-20°C		
79727	3x PL-02 Assay Buffer	50 ml	-20°C	freeze/ thaw	
79728	Blocking Buffer 2	50 ml	+4°C	cycles	
79670	ELISA ECL Substrate A (translucent bottle)	6 ml	Room Temp		
	ELISA ECL Substrate B (brown bottle)	6 ml	Room Temp		
79699	White 96-well microplate	1	Room Temp		

*The concentration of the protein is lot-specific and will be indicated on the tube.

Materials Required but Not Supplied

- PBS (Phosphate Buffer Saline)
- Luminometer or microplate reader capable of reading chemiluminescence
- Rotating or rocker platform

Storage Conditions



This assay kit will perform optimally for up to 6 months from date of receipt when the materials are stored as directed. *Avoid multiple freeze/thaw cycles!*

Safety



This product is for research purposes only and not for human or therapeutic use. This product should be considered hazardous and is harmful by inhalation, in contact with skin, eyes, clothing, and if swallowed. If contact occurs, wash thoroughly.

Contraindications

PCSK9(His)-LDLR Binding Assay Kit is compatible with up to 1% final DMSO concentration. We recommend preparing the inhibitor in no higher than 10% DMSO solution in buffer and using 5 µl per well.

Assay Protocol

- All samples and controls should be performed in duplicates.
- The assay should include a "Blank", "Uncoated control" and a "Positive control".

Coating the plate with LDLR:

 Choose the neutral or acidic buffer at which you wish to test the interaction between PCSK9 and LDLR. You can either run the reaction using acidic PL-01 Assay Buffer (pH 6.0) or neutral PL-02 Assay Buffer (pH 7.4).



2

- 2) Thaw LDLR on ice. Briefly spin the tube containing LDLR to recover the full contents of the tube. If the assay plate is going to be used more than once, calculate the amount of protein required for this portion of the assay (100 ng/well). Aliquot the remaining undiluted protein into single use aliquots depending on how many times the plate will be used and immediately store the aliquots at -80°C.
- 3) Dilute LDLR to 2 ng/ μ l in PBS (enough for 50 μ l/well).
- 4) Add 50 μl of diluted LDLR to each well except "Uncoated Control" and incubate overnight at 4°C. *Do not freeze and reuse the diluted protein.*
- 5) The following day, dilute the 3x Assay Buffer of your choice to 1x Assay Buffer by adding one volume of 3x Assay Buffer to two volumes of distilled water.
- 6) Wash the plate 3 times with 100 μ l/well of 1x Assay Buffer. Tap the plate onto clean paper towels to remove the excess liquid.
- 7) Block by adding 100 μl of Blocking Buffer 2 to each well. Incubate for 1 hour at room temperature. Remove the Blocking Buffer from the plate.

Step 1:

- 1) Prepare the Master Mix: N wells x (10 μ l of 3x Assay Buffer + 15 μ l of distilled water).
- Add 25 μl of Master Mix to each well, including "Blank", "Uncoated control", "Positive Control" and "Test Inhibitor".
- 3) Prepare the Test Inhibitor (5 μ l/well): for a titration, prepare serial dilutions at concentrations 10-fold higher than the desired final concentrations. The final volume of the reaction is 50 μ l.
 - a) If the Test Inhibitor is water-soluble, prepare serial dilutions in the Assay Buffer, 10-fold more concentrated than the desired final concentrations. For the positive and negative controls, use 1x Assay Buffer (Diluent Solution).

OR

b) If the Test inhibitor is soluble in DMSO, prepare the test inhibitor at 100-fold the highest desired concentration in DMSO, then dilute the inhibitor 10-fold in 1x Assay Buffer to prepare the highest concentration of the 10-fold intermediate dilutions. The concentration of DMSO is now 10%.

Prepare serial dilutions of the Test Inhibitor at concentrations 10-fold higher than the desired final concentrations using 10% DMSO in 1x Assay Buffer to keep the concentration of DMSO constant.

For positive and negative controls, prepare 10% DMSO in 1x Assay buffer (vol/vol) so that all wells contain the same amount of DMSO (Diluent Solution).

The final concentration of DMSO should not exceed 1%.



- 4) Add 5 μl of inhibitor solution to each well designated "Test Inhibitor." For the "Positive Control," "Uncoated Control" and "Blank," add 5 μl of Diluent Solution (for example 10% DMSO in 1x Assay Buffer).
- 5) Add 20 µl of 1x Assay Buffer to the well designated "Blank."
- 6) Thaw PCSK9(His) on ice. Briefly spin the tube containing the protein to recover the full contents of the tube. Calculate the amount of protein required for the assay and dilute enough for the assay (50 ng/well).

Aliquot unused, undiluted protein into 2-4 aliquots as may be necessary for future use (single use aliquots) and store them at -80°C.

Note: The protein is sensitive to freeze/thaw cycles. Avoid multiple freeze-thaw cycles. Do not re-use thawed aliquots and do not reuse the diluted protein.

7) Initiate the reaction by adding 20 μl/well of diluted PCSK9(His) to "Uncoated control", "Positive Control" and "Test Inhibitor" (do NOT add to "Blank"). Incubate at room temperature for two hours.

	Blank	Uncoated Control	Positive Control	Test Inhibitor
Master Mix	25 μl	25 μl	25 μl	25 μl
Test Inhibitor	-	-	-	5 μl
Diluent Solution (no Inhibitor)	5 μl	5 μl	5 μl	-
1x Assay Buffer	20 µl	-	-	-
PCSK9(His) (2.5 ng/μl)	-	20 µl	20 µl	20 µl
Total	50 μl	50 μl	50 μl	50 μl

- 8) Wash the plate 3 times with 100 μ l/well of 1x Assay Buffer. Tap the plate onto clean paper towels to remove the excess liquid.
- 9) Block wells by adding 100 μ l of Blocking Buffer 2 to each well. Incubate for 10 minutes at room temperature. Remove the Blocking Buffer from the plate.

Step 2:

- 1) Dilute Anti-His HRP-D antibody 1000-fold with Blocking Buffer 2.
- 2) Add 100 µl to each well. Incubate for 1 hour at room temperature with slow shaking.
- 3) Wash the plate three times with 1x Assay Buffer. Tap the plate onto clean paper towels to remove the excess liquid.
- 4) Block wells by adding 100 μ l of Blocking Buffer 2 to each well. Incubate for 10 minutes at room temperature. Remove the liquid from the plate. Tap the plate onto clean paper towels to remove the liquid.



Step 3:

- Just before use, mix N wells x (50 μl of ELISA ECL substrate A and 50 μl of ELISA ECL substrate B), then add 100 μl of the mix to each well. Discard any unused chemiluminescent mix after use.
- 2) Immediately read in a luminometer or plate reader capable of reading chemiluminescence. The "Blank" value is subtracted from all readings.

Reading Chemiluminescence

Chemiluminescence is the emission of light (luminescence) which results from a chemical reaction. The detection of chemiluminescence requires no wavelength selection because the method used is emission photometry and is not emission spectrophotometry. To properly read chemiluminescence, make sure the plate reader is set for LUMINESCENCE mode. Typical integration time is 1 second, delay after plate movement is 100 msec. Do not use a filter when measuring light emission. Typical settings for the Synergy 2 BioTek plate reader are: use the "hole" position on the filter wheel; Optics position: Top; Read type: endpoint. Sensitivity may be adjusted based on the luminescence of a control assay without enzyme (typically we set this value as 100).

Example Results

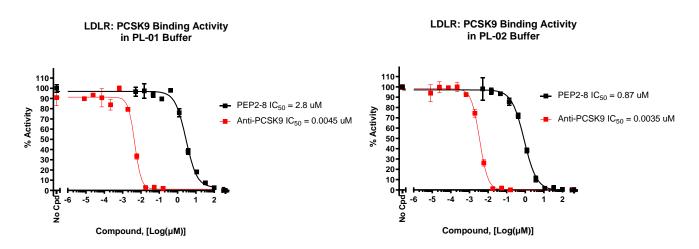


Figure 2: PCSK9-LDLR binding activity in neutral or acidic buffer.

PCSK9-LDLR binding activity was measured using the PCSK9(His)-LDLR Binding Assay Kit (BPS Bioscience #78813) using PL-01 or PL-02 buffers in the presence of increasing concentrations of Anti-PCSK9 Neutralizing Antibody (BPS Bioscience #71207) and PCSK9 inhibitor PEP2-8 (MedChemExpress #HY-P2276). Luminescence was measured using a Bio-Tek microplate reader.

Data shown is representative. For lot-specific information, please contact BPS Bioscience, Inc. at info@bpsbioscience.com

Troubleshooting Guide

Visit bpsbioscience.com/assay-kits-faq for detailed troubleshooting instructions. For all further questions, please email support@bpsbioscience.com



5

Related Products

Products	Catalog #	Size
PCSK9-LDLR Homogeneous Assay Kit	78812	384 reactions
PCSK9, C-terminal His-Avi-Tag, Biotin-Labeled Recombinant	71304	25 μg
LDLR, FLAG-tag Recombinant	71205	50 µg
LDLR, Biotin-Labeled Recombinant	71206	25 μg
Anti-PCSK9 Neutralizing Antibody	71207	50 µg
PCSK9-LDLR TR-FRET Assay Kit	72010	384 reactions
PCSK9(D374T)-LDLR TR-FRET Assay Kit	72011	384 reactions



6